



Standard Test Method for Diagnostic Soil Test for Plant Growth and Food Chain Protection¹

This standard is issued under the fixed designation D 5435; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope*

1.1 This test method covers the determination of quantity (Q) and intensity (I) results for several elements in soils, spoils, fly-ash, and other soil substitutes to ascertain their suitability for the growth of vegetation and possible adverse effects of metals on the food chain.

1.2 The values stated in SI units are to be regarded as the standard.

1.3 *This standard does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 ASTM Standards:

D 653 Terminology Relating to Soil, Rock, and Contained Fluids²

D 3740 Practice for Minimum Requirements for Agencies Engaged in the Testing and/or Inspection of Soil and Rock as Used in Engineering Design and Construction²

E 617 Specification for Laboratory Weights and Precision Mass Standards³

2.2 EPA Standard:

U.S. EPA Method 3050 A²

3. Summary of Test Method

3.1 A representative sample material to be tested is air dried, sieved to pass a 2-mm screen, equilibrated for 16 to 20 h with the soil test solution containing the optimum activities (I) of H_3O^+ , Ca^{++} , Mg^{++} , K^+ and DTPA (diethylenetriaminepentaacetate) to render a small exchange of metals

from the solid phase to the solution phase. This test method is calibrated with respect to pH, K, Ca, Mg, $\text{H}/(\text{Ca} + \text{Mg})^{1/2}$, Mg/K, Ca/Mg, Mn, Fe, Cu, Zn, Al, Pb, Ni, and Cd. In addition to the exchangeable or labile amounts (Q), or both, of the same elements, comparable levels of the negative logarithm values analogous to pH may be calculated using the ligand constants for the respective elements.

4. Significance and Use

4.1 The bioavailability of chemical elements is poorly related to the chemical composition of soils and plant growth media containing a mineral or any type of adsorbed phase. The chemical potential (π for element, i,) is an intensity parameter (I), and the sorbed amount in equilibrium with the soil solution is a measure of the quantity (Q). These parameters for each element (essential or toxic) should be measured in the presence of other elements at or near the desired intensity. This test method is the only method that generates these results simultaneously for several elements. The computer software allows these values to be related to the total sorbed quantities of the different elements. For many substrates, it has been found that the theory for the method holds to the degree that vegetation has been established on many non-soil substrates and soil-water-food chain problems have been evaluated by this test method. This test method has been used on many sites in Pennsylvania and other locations to monitor the effect of sewage sludge applications on land as a source of essential elements for plants with no harmful effects on the food chain. It has also been used to evaluate synthetic soils produced from fly-ash alone or as a component of coal refuse for the establishment of vegetation on mine spoils, coal refuse piles, and abandoned mine lands.

4.2 The quality of the result produced by this standard is dependent on the competence of the personnel performing it, and the suitability of the equipment and facilities used. Agencies that meet the criteria of Practice D 3740 are generally considered capable of competent and objective testing/sampling/inspection/ and the like. Users of this standard are cautioned that compliance with Practice D 3740 does not in

¹ This test method is under the jurisdiction of ASTM Committee D18 on Soil and Rock and is the direct responsibility of Subcommittee D18.22 on Soil as a Medium for Plant Growth.

Current edition approved July 10, 2003. Published July 2003. Originally approved in 1993. Last previous edition approved in 1998 as D 5435-93(1998).

² Available from Superintendent of Documents, U.S. Government Printing Office, Washington, DC 20402.

³ *Annual Book of ASTM Standards*, Vol 14.02.

itself assure reliable results. Reliable results depend on many factors; Practice D 3740 provides a means of evaluating some of those factors.

5. Interferences

5.1 Quality control in reagent preparation is more difficult than for methods that involve fewer elements and compounds. External contamination is a problem to be anticipated. Rubber is a source of Zn and Cd contamination. Rubber gloves used to remove glassware from an acid bath must be removed for subsequent rinse procedures. The blank solution should be tested for Na and other elements. Often the superfloc contains Na which should be removed by use of a cation exchange resin. In addition the filter paper should be checked for Na.

5.2 The cross contamination from glassware is a serious problem. All glassware is soaked in 1 + 1 HNO₃ for 2 h; rinsed with distilled water three times followed with rinsing in distilled, deionized water three times. Remove rubber gloves after first rinsing with distilled water to prevent contamination with Zn.

5.3 The testing solution should be prepared fresh every week. Limited studies indicate that the relative amounts of the different elements desorbed from soils by DTPA change as the prepared solution ages.

5.4 The effect of Ca and other ions on the atomic absorption determination of Cd is significant and requires that an adjustment be made. Background correction will eliminate this source of error.

5.5 The TEA (triethanolamine) is used to prevent excessive dissolution of trace metals from high p-H soils. The buffer capacity is not sufficient, however, to prevent a reflection of soil acidity on trace element availability in soil.

5.6 For some soils the colorimetric determination of phosphorus is not adequate because of the development of turbidity with stannous chloride used as the reducing agent or when *P* is very near the detection limit of 1 µm/L. Other reducing agents as proposed by Watanabe and Olsen (1)⁴ or the isobutyl alcohol extraction method as proposed for water soluble *P* (see section 12.9) have been used with limited success on these samples.

6. Apparatus

6.1 *No. 10 Sieve*, (U.S. Sieve Series Standard 2 mm) constructed from polyethylene.

6.2 *Erlenmeyer Flasks*, 125-mL capacity with polyethylene-covered rubber stoppers or equivalent.

6.3 *Rotating Shaker*, 150 rpm for 125-mL flasks.

6.4 *Filter Funnels*, 11 cm.

6.5 *Filter Paper*, ⁵ 15 cm in diameter.

6.6 *Sample Storage Bottles*, 50 mL polyethylene.

6.7 *pH Meter*, with reference and glass electrodes, that is accurate to ±0.02-pH units when standardized against pH 4.01 and pH 7.00 buffers.

6.8 *Precise Automatic Diluter*, for use with soil test solutions and standards 1:50 in SrCl₂ solution for Mg and Ca determinations by flame AA or ICP.

6.9 *Automatic Diluter*, accurate, calibrated, for diluting soil test solutions that are above the range of the working standards.

6.10 *Atomic Absorption or ICP Spectrometer*, equipped with background correction.

6.11 *Colorimeter*, with 2-cm light-path cells or autoanalyzer for colorimetric analyses.

6.12 *Analytical and Top-Loading Balances*, accuracy to be verified using Specification E 617 weights.

6.13 *50-mL Test Tubes, 15-mL Volumetric Pipettes, and 1-mL Measuring Pipettes*, for use in *P* determinations.

6.14 *50-mL Erlenmeyer Flasks, 10-mL Volumetric Pipettes, and 1-mL Measuring Pipettes*, for *S* determination.

6.15 *Various Beakers*, Erlenmeyer flasks, graduated cylinders, volumetric pipettes, flasks, storage bottles, magnetic heating and stirring unit, refrigerator, and other equipment as required for preparing and storing stock solutions, soil test solutions, reagents, and standards.

6.16 *Computer Facilities*—Computer software that is compatible with IBM-PC and mainframe computers is available to produce the chemical potentials (negative logarithm of the activity of respective ions in soil or substrate solutions) is available. The software is beneficial in producing output that provides decision support for users of this test method.

7. Reagents and Materials

7.1 *Stock Solutions*—Prepare the following stock solutions by dissolving the indicated amount of the pure salt in pure water and diluting to 1 L:

Stock Solution	Amount of Salt per Litre of Solution
0.10 M NaCl	5.84 g of NaCl
0.25 M KCl	18.64 g of KCl
1.00 M MgCl ₂	203.31 g of MgCl ₂ ·6H ₂ O
2.00 M CaCl ₂	294.04 g of CaCl ₂ ·2H ₂ O
0.010 M KH ₂ PO ₄	1.3609 g of KH ₂ PO ₄
1000 mg/l S	5.435 g of K ₂ SO ₄

The use of the above reagents to prepare stock solutions enables a more constant pH to be achieved when TEA is added than the use of compounds considered to provide standards of more accurate composition. Since the chloride salts are not suitable for primary standards, they should be compared with reliable, primary standards and the volumes used are adjusted to meet the required final concentrations.

7.2 *Primary Standards*—Use commercial primary standards for AA or ICP applications, or both, or prepare 1000-mg/L stock solutions of Al, Zn, and Cd by dissolving 1.000 g of the pure metal in 50 mL of 1:1 concentrated HCl-water mixture and diluting to 1 L with pure water. Prepare 1000 mg/L stock solutions of Mn, Fe, Ni, and Cu by dissolving 1.000 g of the pure metal in 1:1 concentrated HNO₃-water mixture and diluting to 1 L with pure water.

7.3 *DTPA*, diethylenetriaminepentaacetic acid.

7.4 *TEA (0.0275 M TEA)*—Dissolve 4.101 g of TEA in pure water and dilute to 1 L.

7.5 *Superfloc-127 Solution, 1 %*—To 700 mL of pure water in a 1-L beaker on a magnetic stirring unit, slowly add 10.0 g of Superfloc-127, a small portion at a time, with continuous stirring and gentle heating. Cover the beaker with a watch glass and allow this solution to stir overnight. Transfer this solution quantitatively to a 1-L volumetric flask and dilute to 1 L. Store this solution in the refrigerator. Dilute 10 mL of the solution to

⁴ The boldface numbers in parentheses refer to a list of references at the end of this test method.

⁵ S & S Number 402S, or equivalent, has been found suitable for this purpose.

100 mL and check for Na and other contaminants. A cation exchange resin can be used to remove the Na.

7.6 Preservative Solution—Dissolve 0.1 g of phenylmercuric acetate in 20 mL of dioxane and dilute to 100 mL with pure water. This solution will be cloudy. Store in the refrigerator.

7.7 Baker Soil Test⁶ Solution—Dissolve 1.5734 g of DTPA in 300 mL of pure water with gentle heating. Transfer this solution quantitatively to a 1-L volumetric flask. Add to this the indicated amounts of the following stock solutions:

Stock Solution	Amount of Each Stock Solution per Litre of Soil Test Solution, mL
0.25 M KCl	10.00
1.00 M MgCl ₂	10.00
2.00 M CaCl ₂	25.00
1 % Superfloc-127 solution	100.00
Preservative solution	10.00

7.7.1 The solution will turn cloudy upon adding the preservative solution. Vigorous mixing will turn the solution clear. Dilute the flask contents to 1 L with pure water and store the soil test solution in the refrigerator. Prepare a fresh test solution weekly.

7.8 SrCl₂ Diluting Solution—Dissolve 161.2 g of SrCl₂·6H₂O in pure water and dilute to 1 L. Dilute 40.0 mL of this solution to 1 L with pure water.

7.9 Ammonium Molybdate Reagent—Dissolve 4.17 g of (NH₄)₆Mo₇O₂₄·4H₂O in 33 mL of pure water. Add to this 47 mL concentrated H₂SO₄. When cool, add 87 mL of pure water. Store in a dark-colored glass bottle in the refrigerator.

7.10 Stannous Chloride Reagent—Dissolve 5 g of SnCl₂·2H₂O in 12.5 mL of concentrated HCl. Store in a dark-colored glass bottle in the refrigerator. Dilute 1 mL of this solution to 75 mL with pure water for use on a daily basis.

7.11 Acid Seed Reagent (6 N HCl containing 20 mg/1 S as K₂SO₄)—Dilute 40.0 mL of the 1000-mg/L S standard to 1 L with pure water. Mix 50.0 mL of this solution with 50.0 mL concentrated HCl to obtain the acid seed reagent.

7.12 BaCl₂·2H₂O Crystals, 20 to 60 mesh material is required.

7.13 Stock Standard Solution—Prepare a combined stock standard by pipetting the indicated amount of each stock solution listed as follows into a 1-L volumetric flask and diluting to volume with pure water:

Stock Solution	Amount of Stock Solution per Litre of Combined Stock Standard, mL	Element Concentration in Combined Stock Standard
0.10 M NaCl	100.0	10 × 10 ⁻³ M
0.25 M KCl	40.0	10 × 10 ⁻³ M
1.00 M MgCl ₂	40.0	40 × 10 ⁻³ M
2.00 M CaCl ₂	100.0	200 × 10 ⁻³ M
1000 mg/L Al	80.0	80 mg/L
1000 mg/L Mn	80.0	80 mg/L
1000 mg/L Fe	80.0	80 mg/L
1000 mg/L Ni	20.0	20 mg/L
1000 mg/L Cu	20.0	20 mg/L
1000 mg/L Zn	30.0	30 mg/L
1000 mg/L Pb	20.0	20 mg/L
1000 mg/L Cd	20.0	20 mg/L

7.14 Compensating Solution—Dissolve 1.573 g of DTPA in 300 mL of pure water with gentle heating. Transfer this solution quantitatively to a 1-L volumetric flask. Add to this 10.0 mL of the preservative solution and 100.0 mL of the 1 % Superfloc-127⁶ solution and dilute to 1 L with pure water. Store in the refrigerator.

7.15 Working Standards for Atomic Absorption Spectrophotometer—Prepare ten flame atomic absorption standards by pipetting the indicated amount of combined stock standard for each working standard listed in Table 1 into a 500-mL volumetric flask, adding 50.0 mL of compensating solution and 5.0 mL of concentrated HNO₃, and diluting to volume with pure water.

7.16 Phosphorus Working Standards—Dilute 100.00 mL of the 0.010 M KH₂PO₄ stock solution to 1 L with pure water to give a solution 0.001 M P. Prepare phosphorus working standards in the range 0 to 20 × 10⁻⁶ M P by diluting aliquots of the 0.001 M from standard to 100 mL with pure water. Prepare these standards for use on a daily basis. Five or six standards should be sufficient to prepare a calibration curve.

TABLE 1 Working Standards for Atomic Absorption Spectrophotometer

Working Standard Number		Amount of Combined Stock Standard per 500 mL of Working Standard				
		1	2	3	4	5
		1	2	3	4	5
		2	4.0			
		3	30.0			
		4	20.0			
		5	12.50			
		6	8.00			
		7	6.00			
		8	4.00			
		9	2.00			
		10	1.00			
Elemental Concentrations of Working Standards						
Element	Unit	Standard Number				
		1	2	3	4	5
Na	10 ⁻⁴ M	10.0	8.0	6.0	4.0	2.5
K	10 ⁻⁴ M	10.0	8.0	6.0	4.0	2.5
Mg	10 ⁻⁴ M	40.0	32.0	24.0	16.0	10.0
Ca	10 ⁻⁴ M	200.0	160.0	120.0	80.0	50.0
Al	mg/L	8.0	6.4	4.8	3.2	2.0
Mn	mg/L	8.0	6.4	4.8	3.2	2.0
Fe	mg/L	8.0	6.4	4.8	3.2	2.0
Ni	mg/L	2.0	1.6	1.2	0.8	0.5
Cu	mg/L	2.0	1.6	1.2	0.8	0.5
Zn	mg/L	3.0	2.4	1.8	1.2	0.75
Pb	mg/L	2.0	1.6	1.2	0.8	0.5
Cd	mg/L	2.0	1.6	1.2	0.8	0.5
Element	Unit	Standard Number				
		6	7	8	9	10
Na	10 ⁻⁴ M	1.6	1.2	0.8	0.4	0.2
K	10 ⁻⁴ M	1.6	1.2	0.8	0.4	0.2
Mg	10 ⁻⁴ M	6.4	4.8	3.2	1.6	0.8
Ca	10 ⁻⁴ M	32.0	24.0	16.0	8.0	4.0
Al	mg/L	1.28	0.96	0.64	0.32	0.16
Mn	mg/L	1.28	0.96	0.64	0.32	0.16
Fe	mg/L	1.28	0.96	0.64	0.32	0.16
Ni	mg/L	0.32	0.24	0.16	0.08	0.04
Cu	mg/L	0.32	0.24	0.16	0.08	0.04
Zn	mg/L	0.48	0.36	0.24	0.12	0.06
Pb	mg/L	0.32	0.24	0.16	0.08	0.04
Cd	mg/L	0.32	0.24	0.16	0.08	0.04

⁶ Baker Soil Test is a registered trademark of Land Management Decisions to protect the name and enable the software to be maintained and updated as required to provide “users” of franchised laboratories with maximum benefits from this test method.

7.17 Sulfur Working Standards—To 100.0 mL of the 1000 mg/L sulfur standard, add 100.0 mL of the soil test solution and dilute to 1 L with pure water to yield a 100 mg/L sulfur standard. Prepare the working sulfur standards by pipetting the indicated amounts of 100 mg/L sulfur standard and soil test solution for each sulfur standard listed as follows into a 100-mL volumetric flask and diluting to volume with pure water. Prepare these standards for use on a daily basis:

Standard Number	Amount of 100 mg/L S Standard per 100 mL of Working S Standard, mL	Amount of Soil Test Solution per 100 mL of Working S Standard, mL	S Concentration in Working S Standard, mg/L
1	0.00	10.00	0
2	2.00	9.80	2
3	5.00	9.50	5
4	10.00	9.00	10
5	20.00	8.00	20
6	30.00	7.00	30
7	40.00	6.00	40

8. Sample Preparation

8.1 Sample Preparation—Air dry and crush the soil or soil substitute sample to pass a 2-mm sieve.

8.2 Synthetic Soil Mixtures—For mixtures of fly-ash, spoils, compost, etc. to make a synthetic soil, the materials should be prepared as soil and the mixture should be wetted, air-dried, and mixed at least two and preferably three times before it is tested. Otherwise, the system will be changing and the test solution cannot reach equilibrium.

9. Calibration

9.1 Working Standards—Chose working standards from those described in 7.11. Dilute samples with element concentrations above the range of the instrument or standards with solution containing 100 mL of compensating solution per litre.

9.2 Calibration—Use the working standards to prepare the necessary calibration curves for calculating the element concentrations in the samples. The number of standards used will depend on the linear range for each instrumental method. Pay careful attention to the instrument manufacturer's instruction in this regard. Computers greatly aid in the preparation of calibration curves, error analysis, and quality-control programs.

10. Procedure

10.1 Blank Preparation—Prepare a blank soil test solution by pipetting 40.0 mL of pure water (RO + deionized or equivalent), 5.0 mL of soil test solution, and 5 mL of 0.0275 M TEA into a 125-mL Erlenmeyer flask and mix. Check the pH of this solution. It should be 7.3 ± 0.05 ; if not, adjust the amount of 0.0275 M TEA and pure water used to obtain a pH of 7.3 ± 0.05 in a final volume of 50.0 mL of soil test solution. Greater precision can be achieved by working with 1-L volumes for 20 samples at a time.

10.2 Baker Soil Test Equilibration—Weigh 5.000 g of soil (< 2 mm) into a 125-mL Erlenmeyer flask. Add to the flask the amount of pure water needed to obtain a pH of 7.3 ± 0.05 and a final volume of 50.0 mL with no soil present as determined in the blank preparation step (see 10.1). Add 5.0 mL of soil test solution to the flask. Add to the flask the amount of 0.0275 M TEA needed to obtain a pH of 7.3 ± 0.05 and a final volume

of 50 mL with no soil present as determined in the blank preparation step (see 10.1). Stopper the blank and sample flasks with plastic stoppers and place on a rotating shaker for 1 h at 150 oscillations per minute. Allow the flasks to stand for an additional 21 to 23 h and filter the samples through filter paper.⁵

10.3 pH—Determine the pH of an aliquot of sample using a pH meter and glass and reference electrodes standardized against pH 4.01 and 7.0 buffers.

10.4 Elemental Analysis—The actual analysis of sample will vary with instrumentation. Na and K may be analyzed by flame emission spectroscopy; for Ca and Mg by Flame AA, the test solution and standards may need to be diluted 1:50 in the SrCl₂ diluting solution using a precise automatic diluter. Both Ca and Mg may be determined by Flame AA. All other elements may be determined by Flame or Flameless AA, or both, depending on the concentrations in solution as well as by ICP.

10.5 Na and K—These two elements are easily determined in the blank and soil test solution samples by flame emission spectroscopy.

10.6 Mg and Ca—Dilute the standards and blank and soil test solutions samples 1:50 in the SrCl₂ diluting solution using a precise automatic diluter. Determine the concentrations of these elements in the blank and samples by atomic absorption spectrophotometry or by ICP.

10.7 Al, Mn, Fe, Ni, Cu, Zn, and Cd—Determine the concentration of these elements in the blank and soil test solution samples by atomic absorption spectrophotometry.

10.8 P—Add 0.5 mL of the ammonium molybdate reagent to 15.00 mL each of blank and soil test solution samples and each P standard, and mix. Add 0.3 mL of the stannous chloride reagent, mix, and read the absorbencies on a colorimeter at 660-mu using a cell with a 2-cm light path after 6 but before 11 min.

10.9 Sulfate, S—Add 1 mL of acid seed reagent to 10.00 mL of blank and sample soil test solutions and each S standard, mix, and add 0.5 g of BaCl₂·2H₂O crystals (20 to 60 mesh). Let stand for exactly 1 min, mix until the crystals are dissolved, and read the absorbencies on a colorimeter at 420 mu between 2 and 8 min using a cell with a 2-cm light path.

11. Calculation

11.1 Report the results in the units of the standards for P, K, Ca, and Mg. For the trace elements, report the soil levels are reported (solution levels: $\times 10.0$). If filtrates are diluted, the dilution factor must be applied.

11.2 Computer programs are available to perform the iterative calculations required to produce values for pH, pK, pCa, pMg, pFe, pMn, etc. The validity of these conversions has been verified using numerous samples from research investigations and thousands of user samples. Use of this computerized expert system provides an intensity value, and a desorbed quantity amount for most elements and relative intensity values for K, Ca, Mg, and H. In addition, data for Total Sorbed Metals by the

U.S. Environmental Protection Agency (EPA) method is included. See Footnote ⁷ for information regarding the computer program. The iterative procedure may also be performed using program GEOCHEM-PC (2, 3).⁷ See Footnote 10 for information regarding the computer program to provide the decision support output from the database for the method.

12. Interpretation of Results

12.1 For those that do not have access to the computer programs Table 2 shows interpretation offered with respect to plant growth and food-chain protection.

12.2 As indicated in 11.2, the Baker Soil Test⁸ software allows the inclusion of total sorbed metals by U.S. EPA Method 3050A. The above interpretative data represent “small exchange” calibration values from the existing data for over 10 000 samples. The numbers given here do not refer to total metal content of a soil sample or the results by U.S. EPA Method 3050A, and should not be compared with previously published ranges of soil element concentrations. Critical levels for the quantity values presented above, based on field re-

sponse data, varies with crops and local field conditions. The low levels are not usually deficient and high levels are not usually toxic. The availability (plant response) is predicted more accurately by the respective p_i values as calculated by Baker Soil Test⁸ computer software. Equivalent p_i values can be obtained by use of the computer program GEOCHEM⁹. Relative values for the three parameters quantity (above), intensity ($-\log a_i$), and total sorbed metals provide an evaluation of a particular soil against the data-base summarized by Baker, Lotse, and Amistadi (4).⁹

13. Effects of Storage

13.1 Soil samples may be stored in an air-dry condition for prolonged periods with no effect on test levels of the different elements. However, maintaining samples in a wet condition at room temperature renders the results for Mn, Fe, Cu, P and possibly other elements unreliable. Differences between moist and air-dried samples have been found, especially for Mn and Al.

14. Precision and Bias

14.1 *Precision*—The within laboratory standard deviations and maximum allowable differences are as follows:

Element	Standard Deviation	Maximum Difference
K	0.088	0.23
Ca	1.691	4.40
Mg	0.661	1.72
Mn	2.184	5.68
Fe	0.176	0.458
Cu	0.016	0.042
Zn	0.042	0.109
Na	0.266	0.692
Al	0.096	0.250
Ni	0.021	0.055
Pb	0.039	0.101
Cd	0.042	0.109

14.2 *Bias*—There is no acceptable reference value for this test method; therefore, bias cannot be determined.

15. Keywords

15.1 diagnostic soil test; food chain; plant

⁷ GEOCHEM is a computer program published by Sposito and Mattigod.

⁸ Information may be obtained from Land Management Decisions, Inc., 1429 Harris Street, State College, PA 16803.

TABLE 2 Interpretation of Results

Element ^A	Low	Normal	High
P $10^{-4}M$	1	1 to 2.5	2.5 to 5
K $10^{-4}M$	1.5	1.5 to 4	4 to 8
Mg $10^{-4}M$	5	5 to 16	16 to 20
Ca $10^{-4}M$	40	40 to 60	60 to 120
Al mg/kg	5	5 to 12.5	12.5 to 20
Mn mg/kg	20	20 to 50	50 to 100
Fe mg/kg	10	10 to 50	50 to 75
Cu mg/kg	2	2 to 10	10 to 30
Zn mg/kg	2	2 to 40	40 to 70
Pb mg/kg	2	2 to 10	10 to 30
Cd mg/kg	0.1	0.1 to 0.85	0.85 to 1.6
Na mg/kg	50	50 to 100	100 to 150
S mg/kg	20	20 to 50	50 to 100
Ni mg/kg	0.5	0.5 to 5	5 to 15
Mo mg/kg	0.04	0.04 to 0.5	0.5 to 5

^A P, K, Ca, and Mg are expressed as concentrations in the equilibrating solution and all others are amounts desorbed or solubilized from the soil (solution concentrations $\times 10$).

REFERENCES

- (1) Watanabe, F. S., and Olsen, S. R., “Colorimetric Determination of Phosphorus in Water Extracts of Soil,” *Soil Sci.* 93, 1962, pp. 183–188.
- (2) Sposito, G., and Mattigod, S. V., *GEOCHEM: A Computer Program for the Calculation of Chemical Equilibria in Soil Solutions and Other Natural Water Systems*, Kearney Foundation of Soil Science, Univ. of CA, Riverside, 1980.
- (3) Stout, W. L., and Baker, D. E., “A New Approach to Soil Testing: III. Differential Adsorption of Potassium,” *Soil Sci. Soc. Am. J.* 42, 1978, pp. 307–310.
- (4) Baker, D. E., Lotse, E., and Amistadi, M. K., “Testing Soils for Metals With Emphasis on Cadmium in the Food Chain,” *Proceedings, ASTM Conf. on Application of Agri. Anal. in Environ. Studies*, Atlantic City, N.J., 1991, (In Press).
- (5) Baker, D. E., “A New Approach to Soil Testing,” *Soil Sci.* 112, 1971, pp. 381–391.
- (6) Baker, D. E., “A New Approach to Soil Testing: II. Ionic Equilibria Involving H, K, Ca, Mg, Mn, Fe, Cu, Zn, Na, P and S,” *Soil Sci. Soc. Am. Proc.* 37, 1973, pp. 537–541.

- (7) Baker, D. E., "Soil Chemical Constraints in Tailoring Plants to Fit Problem Soils I, Acid Soils," *Plant Adaptation to Mineral Stress in Problem Soils*, M. J. Wright (Ed.), Cornell University, Ithaca, N.Y. 14853, 1976.
- (8) Baker, D. E., "Ion Activities and Ratios in Relation to Corrective Treatments of Soils," *Soil Testing: Correlating and Interpreting the Analytical Results*, Am. Soc. Agron., Madison, Wis., 1977, pp. 55–74.
- (9) Baker, D. E., "Baker Method," *Handbook on Reference Methods for Soil Testing. Council on Soil Testing and Plant Analysis*, Athens, GA 30602, 1980.
- (10) Baker, D. E. (ed.), "Criteria and Recommendations for Land Application of Sludges in the Northeast," *PA. Agri. Exp. Sta. Bul. 851*, 1985.
- (11) Baker, D. E., "Copper," Chapter 8, *Heavy Metals in Soils*, B. J. Alloway (Ed.), Blackie and Son Ltd., Glasgow and London, pp. 151–176.
- (12) Baker, D. E., Baker "Soil Test Theory and Applications," *Proc. Int. Conf. Soil Testing and Plant Analysis. Comm. Soil Sci. and Plant Anal. 21*, 1990, pp. 981–1008.
- (13) Baker, D. E., and Amacher, M. C., "Development and Interpretation of a Diagnostic Soil Testing Program," *Bul. No. 826, PA. Agri. Exp. Sta.*, University Park, PA, 1981.
- (14) Baker, D. E., Amacher, M. C., and Leach, R. M., "Sewage Sludge as a Source of Cadmium in Soil-Plant-Animal Systems," *Environ. Health Perspectives 28*, 1979, pp. 45–49.
- (15) Baker, D. E., and Bowers, M. E., "Human Health Effects of Cadmium Predicted from Growth and Composition of Lettuce Grown in Gardens Contaminated by Emissions from Zinc Smelters," *Trace Substances in Environmental Health, XXII*, 1988, pp. 281–295.
- (16) Baker, D. E., and Buck, J. K., "Using Computerized Expert Systems, Unique Soil Testing Methods and Monitoring Data in Land Management Decisions," *Mine Drainage and Surface Mine Reclamation Vol II, Mine Reclamation, Abandoned Mine Lands and Policy Issues, Bur. Mines Cir. 9184*, 1988, pp. 246–256.
- (17) Baker, D. E., and Low, P. F., "Effect of the Sol-Gel Transformation in Clay-Water Systems on Biological Activity: II, Sodium Uptake by Corn Seedlings," *Soil Sci. Soc. Am. Proc. 34*, 1970, pp. 49–56.
- (18) Baker, D. E., Rasmussen, D. S., and Kotuby, J., "Trace Metal Interactions Affecting Soil Loading Capacities for Cadmium," *Hazardous and Industrial Waste Management and Testing: Third Symposium, ASTM STP 851*, Larry P. Jackson, Alan R. Rohlik, and Richard A. Conway (Eds.), ASTM, Philadelphia, PA, 1984, pp. 118–132.
- (19) Baker, D. E., and Wolf, A. M., "Soil Chemistry, Soil Mineralogy and the Disposal of Solid Wastes," S. K. Majumdar and E. W. Miller, (Eds.), *Solid and Liquid Wastes: Management, Methods and Socio-economic Considerations*, Pennsylvania Academy of Science, Philadelphia, 1984.
- (20) Hornick, S. B., Baker, D. E., and Guss, S. B., "Crop Production and Animal Health Problems Associated With High Soil Molybdenum," W. R. Chappel and K. K. Petersen (Eds.), *Molybdenum in the Environment, Volume 2, Chapter 11*, 1976, pp. 665–684.
- (21) Leach, R. M., Jr., Wei-Li Wang, Kathy, and Baker, D. E., "Cadmium and the Food Chain: The Effect of Dietary Cadmium on Tissue Composition in Chicks and Laying Hens," *J. Nutr. 109*(3), 1979, pp. 437–443.
- (22) Risser, J. A., and Baker, D. E., "Testing Soils for Toxic Metals," R. L. Westerman (Ed.), *Soil Testing and Plant Analysis, Third Edition*, 1990, pp. 275–298.
- (23) Senft, J. P., and Baker, D. E., "Utilization of the Baker Soil Test in Synthetic Soil Preparation for Reclamation of Coal Ash Disposal Sites," *Proceedings, ASTM Conf. on Application. of Agri. Anal. in Environ. Studies*, Atlantic City, N.J., ASTM, 1991. (In Press)
- (24) Stout, W. L., and Baker, D. E., "Effect of Differential Adsorption of Potassium and Magnesium in Soils on Magnesium Uptake by Corn," *Soil Sci. Soc. Am. J. 45*, 1981, pp. 996–997.
- (25) Watanabe, F. S., and Olsen, S. R., "Test of an Ascorbic Acid Method for Determining Phosphorus in Water and NaHCO₃ Extracts from Soil," *Soil Sci. Soc. Am. Proc. 29*, 1965, pp. 677–678.

SUMMARY OF CHANGES

In accordance with Committee D18 policy, this section identifies the location of changes to this standard since the last edition (93–(1998)) that may impact the use of this standard.

- | | |
|---|--|
| (1) Terminology D 653 and Practice D 3740 were added to the Referenced Documents Section. | (2) 4.2 was added to the Significance and Use Section. |
|---|--|

ASTM International takes no position respecting the validity of any patent rights asserted in connection with any item mentioned in this standard. Users of this standard are expressly advised that determination of the validity of any such patent rights, and the risk of infringement of such rights, are entirely their own responsibility.

This standard is subject to revision at any time by the responsible technical committee and must be reviewed every five years and if not revised, either reapproved or withdrawn. Your comments are invited either for revision of this standard or for additional standards and should be addressed to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee, which you may attend. If you feel that your comments have not received a fair hearing you should make your views known to the ASTM Committee on Standards, at the address shown below.

This standard is copyrighted by ASTM International, 100 Barr Harbor Drive, PO Box C700, West Conshohocken, PA 19428-2959, United States. Individual reprints (single or multiple copies) of this standard may be obtained by contacting ASTM at the above address or at 610-832-9585 (phone), 610-832-9555 (fax), or service@astm.org (e-mail); or through the ASTM website (www.astm.org).